Structural remodelling in AS

Tumour necrosis factor blockers and structural remodelling in ankylosing spondylitis: what is reality and what is fiction?

Georg Schett, Robert Landewé, Désirée van der Heijde

A delineation of the differences in pathology between AS and RA

hallmark of spondylarthropathies (SpA), such as ankylosing spondylitis, (AS) is the fusion of joints as well as intervertebral spaces. This fusion is caused by the formation of bony spurs appearing as syndesmophytes and osteophytes in the intervertebral spaces and in the joints, respectively. Fusion of joints is based on increased endochondral ossification, which allows bone formation and bridges the joint space. Tumour necrosis factor (TNF) is a key proinflammatory cytokine in AS, but is a potent inhibitor of bone formation, and so is unlikely to explain the formation of osteophytes in AS. This is also suggested by recent clinical data showing that TNF blockade seems not to affect structural remodeling of the spinal skeleton in AS, which largely indicates changes due to increased bone apposition. Thus, molecular concepts of structural remodelling in AS need revision, and new pathways involved in bone formation, such as Wingless proteins or transforming growth factor β , might be a clue to the pathogenesis of structural remodelling in AS. The efficacy of TNF blockers to improve clinical symptoms in AS, their poor effect on structural remodelling, and the weak relationship between clinical symptoms and structural damage in AS will profoundly revise our picture of AS in the future.

MECHANISMS OF JOINT FORMATION – MOLECULAR LESSONS FOR JOINT FUSION

Joints and intervertebral spaces form gaps between bones, which allow motion and flexibility. These gaps are actively formed during early development, when chondrogenic formations of the vertebral column and limbs start to branch and build segments. Formation of these gaps depends on the expression of proteins involved in mesenchymal cell differentiation, such as cartilage-derived morphogenic protein 1 (also called GDF5) and bone morphogenic protein (BMP) 5.¹ Without these proteins no joints are formed, since the appropriate differentiation of cells, which form the synovial membrane, are then lacking. Wingless (Wnt) proteins, such as Wnt-14 (also known as Wnt-9a), are also crucial for the initiation of joint formation in the limbs.² Joint formation can thus be considered as an active differentiation process, which replaces the chondrogenic matrix by specific fibroblast-like cells that form the synovial membrane, the periosteum and the joint capsule.

BONY PROTRUSION AS STRESS RESPONSE OF THE JOINT

Joints allow maintaining motion, which, however, requires a structurally intact joint space for smooth gliding of articular surfaces. Inflammation leads to joint damage, which causes pain, swelling, stiffness and functional impairment in patients with chronic inflammatory and degenerative joint disease. Resident mesenchymal tissue in joints, however, is not inert when exposed to an inflammatory attack, and causes certain response patterns, which allow structural remodelling to cope with unphysiological stress. The most prominent pattern is osteophyte formation, which includes spondylophyte and syndesmophyte formation when these structures are located in the axial skeleton. Osteophytes, spondylophytes and syndesmophytes are bony protrusions, which appear on plain radiographs, CT scans and MRI of patients with seronegative SpA, in particular AS, and osteoarthritis (OA), but are virtually absent in rheumatoid arthritis (RA). Syndesmophytes, vertical bony spurs, ultimately leading to a bridge between vertebrae, are a hallmark of AS. Similar lesions, now more horizontally oriented, are also found in degenerative joint diseases such as OA, psoriatic arthritis or haemochromatosis arthropathy, both among vertebral bodies (spondylophytes) and at peripheral joints (osteophytes).

Bony protrusions are based on endochondral ossification, which leads to deposition of the chondrogenic matrix and later to remodelling into bone. Bony spurs emerge from the periosteum close to joints or intervertebral spaces, where mesenchymal cells are localised, which have the ability to differentiate into cartilage and bone, when they receive the appropriate signals. Emergence of osteophytes depends on stress on the joint, and apparently both mechanical stress (as evident from the abundance of such lesions in OA) and inflammatory stress can precipitate their formation. From a pathophysiological point of view these lesions can be seen as an attempt of repair or stabilisation mechanism to reduce motion in the affected joint. Bony spurs can even bridge joints leading to bone ankylosis and complete stabilisation of joints. Longstanding sacroiliitis is a typical example, which, after complete ankylosis and immobilisation of the joint, leads to a marked reduction in clinical symptoms. Bridging syndesmophytes in AS is another clear example.

DIFFERENCES IN INFLAMMATORY BONE REMODELLING BETWEEN RA AND AS

In contrast to AS, RA is the prototype of a disease, which is not associated with osteophyte formation despite severe joint damage (fig 1). The pathophysiological picture of RA is characterised by osteoclast formation and bone destruction, with no or mild signs of bone repair.³ This is based on the dominance of bone resorption in RA, which rapidly destroys the periosteal lining and invades the bone. This process is fuelled by rapid generation of osteoclasts through TNF and receptor activator for nuclear factor κ B ligand (RANKL), and enhanced bone resorption combined with a blunted response of bone formation, which involves inhibitors of Wnt proteins, such as Dickkopf-1 (DKK-1).⁵⁻⁷ The activating role of TNF in osteoclast formation has been defined in the past 5 years, whereas the role of TNF in decreasing osteoblast formation is known for many years but its molecular regulation had been poorly defined until recently.8 9 RA combines rapid bone resorption with inhibition of bone formation leading to unfavourable imbalance of skeletal homeostasis, leading to rapid development of erosions. Structural damage in RA at least partly mimics bone damage of multiple myeloma, since both diseases are characterised by "holes" in the bone with no major reparative response. Importantly, the structural damage measured by radiological scores in RA is a direct consequence of the inflammatory process,

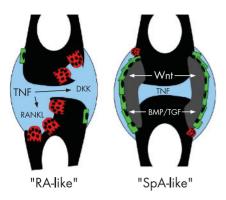


Figure 1 In rheumatoid arthritis (RA), joints undergo progressive resorption, which is mediated by the generation of osteoclasts (red) in the joint. These cells resorb bone and are induced by receptor activator for nuclear factor κ B ligand (ŔANKĹ), which is activated upon tumour necrosis factor (TNF) challenge. Moreover, osteoblasts (green), the bone-forming cells, are suppressed in RA, which is at least partly mediated by Dickkopf (DKK) proteins. In spondylarthopathies (SpA) such as ankylosing spondylitis, bridging of joint and intervertebral spaces by bony spurs (grey) is observed. These changes are based on endochondral ossification and may represent a kind of repair strategy of the joint. TNF drives inflammation in SpA similar to that in RA, but there is no connection to increased bone formation driven by osteoblasts (green), which is based on increased activation of Wingless (Wnt)- proteins, bone morphogenic proteins (BMPs) and transforming growth factor beta (TGF β). Wnt activation also blocks osteoclast differentiation through increased production of osteoprotegerin.

which makes the good relationship between clinical disease activity and structural damage in this disease understandable.¹⁰

By contrast, the pathological, and partly also the radiographic, picture of SpA is dominated by response to stress. Despite chronic inflammation, which is a well-known precipitator of trabecular bone loss and also affects patients with AS, there is evidence for a dramatic bone formation in the periosteal compartment in AS but not in RA. It seems that in AS an initial bone-resorptive phase may act as a stress factor, which is followed by profound endochondral bone formation originating from the periosteum and leading to bone spurs that could bridge the joints and fuse the vertebrae. Molecularly these lesions depend on increased bone formation, which is most likely governed by members of the TGF/ BMP protein family, as well as by the group of Wnt proteins. Thus, intracellular regulators of TGF-B/BMPs, such as regulatory Smads (Smad6, Smad7), block osteophyte formation in murine joints.¹¹ Added to this, Wnt proteins seem to be regulators of osteophyte formation. Binding of Wnt to its receptor Lrp5/6 and activation of downstream regulators such as B-catenin are crucial for bone formation.¹² Moreover, there is a cross-talk with the RANKL-osteoprotegerin system, since Wnt signalling activates osteoprotegerin, which balances RANKL-induced osteoclast activation.13 Inhibitors of Wnt, such as DKK-1, are key target genes of TNF, which probably explains the negative effects of TNF and other proinflammatory cvtokines on bone formation. In line with this, DKK-1 levels are low in AS but high in RA, suggesting that the Wnt signalling cascades are turned on in the joints of AS.⁷ These findings suggest that early developmental programmes are switched on when joints form osteophytes to bridge and stabilise the diseased joint space. It also suggests that TNF negatively regulates bone formation, and that inhibition of bone formation and osteophyte growth by TNFblocking agents is unlikely.

TNF BLOCKERS AND INFLAMMATORY BONE DESTRUCTION

According to the pathophysiological process explained above, it can be expected that drugs that block the action of TNF are helpful in reducing the formation of new erosions in RA, but that their role in reducing structural damage in AS might be fundamentally different. There is abundant information that the available TNF blockers, etanercept, infliximab and adalimumab, can reduce progression of structural damage and may even lead to repair in RA.¹⁴⁻¹⁶ Moreover, in RA, TNF blockers induce a marked reduction in clinical disease activity, which parallels the reduction in structural damage, but there is also reduction of radiographic progression even in patients with some persistent disease activity, especially if a TNF blocker is used in combination with methotrexate.17 As already indicated above, there is a clear longitudinal relationship between clinical disease activity and subsequent radiographic damage in patients with RA,^{10 18} as well as a distinct relationship between inflammation on MRI and subsequent structural damage on both MRI and radiographs.19

TNF BLOCKERS AND BONE APPOSITION IN AS

What is known about the relationship between clinical disease activity, findings on MRI and structural damage on radiographs in AS? Very few studies have been published on this topic. From the Outcome in Ankylosing Spondylitis International Study (OASIS) cohort, we know that radiographic progression is unrelated to clinical disease activity parameters and acute-phase reactants; only MMP3 was an independent predictor of radiographic progression in this cohort.²⁰ Other (indirect) data are available from trials using TNF blockers in AS. There is very good efficacy on clinical disease activity, acute-phase reactants and inflammation visible on MRI.^{21–23} So far there is only one controlled study (published as abstract) addressing the efficacy of a TNF blocker (etanercept) on progression of structural damage.²⁴ Radiographs from the OASIS cohort were used as a comparator, and there was no difference in radiographic progression over a 2-year time frame between patients from OASIS (without a TNF blocker) and patients treated with etanercept. Possible explanations are differences in disease activity and severity between patients from OASIS and patients included in the trial. However, limiting the analysis to patients from OASIS who would have fulfilled the entry criteria for the trial yielded results that were completely comparable. Also, adjustment for all possible differences in baseline variables did not change the results. Another possibility brought forward was that patients already had established disease, and that reduction in radiographic progression could be seen only during the early phases of the disease. Insufficient duration of follow-up was also considered, but seemed unlikely, as 2 years is already a long duration of follow-up and the clinical efficacy is already seen within 2 weeks. Finally, it was considered whether there might be a difference between etanercept and the two anti-TNF antibodies, infliximab and adalimumab, as there is also a difference in efficacy on inflammatory bowel disease between these agents. However, the above-presented data on pathophysiological processes in bone formation, the major abnormality in AS, give the most likely explanation that it cannot be expected that TNF blockers are effective in reducing new bone formation. Data with longer follow-up, with the antibodies against TNF and in patients with early disease, will give us the final answer whether TNF blockers indeed do not inhibit syndesmophyte formation in AS.

Ann Rheum Dis 2007;**66**:709–711. doi: 10.1136/ard.2007.071159

Authors' affiliations

Georg Schett, Department of Internal Medicine 3, Institute for Clinical Immunology, University of Erlangen-Nurnberg, Erlangen, Germany Robert Landewé, Department of Internal Medicine & Rheumatology, University Hospital Maastricht, Maastricht, The Netherlands Désirée van der Heijde, Department of Rheumatology, Leiden University Medical Centre, Leiden, The Netherlands

Correspondence to: Professor G Schett, Department of Internal Medicine 3, Institute for Clinical Immunology, University of Erlangen-Nuernberg, Krankenhausstrasse 12, D-91054 Erlangen, Germany; georg.schett@med3.imed.uni-erlangen.de

Accepted 14 March 2007

Competing interests: None declared.

REFERENCES

- Storm EE, Kingsley DM. Joint patterning defects caused by single and double mutations in members of the bone morphogenetic protein (BMP) family. *Development* 1996;122:3969–79.
- 2 Hartmann C, Tabin CJ. Wnt-14 plays a pivotal role in inducing synovial joint formation in the developing appendicular skeleton. *Cell* 2001;104:341–51.
- 3 Gravallese EM, Harada Y, Wang JT, Gorn AH, Thornhill TS, Goldring SR. Identification of cell types responsible for bone resorption in rheumatoid arthritis and juvenile rheumatoid arthritis. *Am J Pathol* 1998;152:943–51.
- 4 Schett G, Hayer S, Zwerina J, Redlich K, Smolen JS. Mechanisms of disease: the link between RANKL and arthritic bone disease. Nat Clin Pract Rheumatol 2005;1:47–54.
- 5 Shigeyama Y, Pap T, Kunzler P, Simmen BR, Gay RE, Gay S. Expression of osteoclast differentiation factor in rheumatoid arthritis. Arthritis Rheum 2000;43:2523–30.
- 6 Redlich K, Hayer S, Maier A, Dunstan CR, Tohidast-Akrad M, Lang S, et al. Tumor necrosis factor amediated joint destruction is inhibited by targeting osteoclasts with osteoprotegerin. Arthritis Rheum 2002;46:785–92.
- 7 Diarra D, Stolina M, Polzer K, Zwerina J, Ominsky MS, Dwyer D, et al. Dickkopf-1 is a master regulator of joint remodeling. Nat Med 2007;13:156–63.
- 8 Lam J, Takeshita S, Barker JE, Kanagawa O, Ross FP, Teitelbaum SL. TNF-alpha induces osteoclastogenesis by direct stimulation of macrophages exposed to permissive levels of RANK ligand. J Clin Invest 2000;106:1481–8.

- 9 Bertolini DR, Nedwin GE, Bringman TS, Smith DD, Mundy GR. Stimulation of bone resorption and inhibition of bone formation in vitro by human tumour necrosis factors. Nature 1986;319:516.
- Welsing PM, Landewé RB, van Riel PL, Boers M, van Gestel AM, van der Linden S, et al. The relationship between disease activity and radiologic progression in patients with rheumatoid arthritis: a longitudinal analysis. Arthritis Rheum 2004;50:2082–93.
 Scharstuhl A, Vitters EL, van der Kraan PM, van den
- 11 Scharstuhl A, Vitters EL, van der Kraan PM, van den Berg WB. Reduction of osteophyte formation and synovial thickening by adenoviral overexpression of transforming growth factor beta/bone morphogenetic protein inhibitors during experimental osteoarthritis. Arthritis Rheum 2003;48:3442–51.
- Miller JR. The Writs. Genome Biol 2001;3:3001–11.
- 13 Glass DA, Bialek P, Ahn JD, Starbuck M, Patel MS, Clevers H, et al. Canonical Wnt signaling in differentiated osteoblasts controls osteoclast differentiation. Dev Cell 2005;8:751–64.
- 14 Klareskog L, van der Heijde D, de Jager JP, Gough A, Kalden J, Malaise M, et al. Therapeutic effect of the combination of etanercept and methotrexate compared with each treatment alone in patients with rheumatoid arthritis: double-blind randomised controlled trial. *Lancet* 2004;363:675–81.
- 15 Lipsky PE, van der Heijde DM, St Clair EW, Furst DE, Breedveld FC, Kalden JR, et al. Infliximab and methotrexate in the treatment of rheumatoid arthritis. Anti-Tumor Necrosis Factor Trial in Rheumatoid Arthritis with Concomitant Therapy Study Group. N Engl J Med 2000;343:1594–602.
- Study Group. N Engl J Med 2000;343:1594–602.
 Keystone EC, Kavanaugh AF, Sharp JT, Tannenbaum H, Hua Y, Teoh LS, et al. Radiographic, clinical, and functional outcomes of treatment with adalimumab (a human anti-tumor necrosis factor monoclonal antibody) in patients with active rheumatoid arthritis receiving concomitant methotrexate therapy: a randomized, placebo-controlled, 52-week trial. Arthritis Rheum 2004;50:1400–11.
- 17 Landewé R, van der Heijde D, Klareskog L, van Vollenhoven R, Fatenejad S. Disconnect between inflammation and joint destruction after treatment

with etanercept plus methotrexate: results from the trial of etanercept and methotrexate with radiographic and patient outcomes. *Arthritis Rheum* 2006;**54**:3119–25.

- 18 Boers M, Kostense PJ, Verhoeven AC, van der Linden S. Inflammation and damage in an individual joint predict further damage in that joint in patients with early rheumatoid arthritis. Arthritis Rheum 2001;44:2242–6.
- 19 Ostergaard M, Duer A, Nielsen H, Johansen JS, Narvestad E, Ejbjerg BJ, et al. Magnetic resonance imaging for accelerated assessment of drug effect and prediction of subsequent radiographic progression in rheumatoid arthritis: a study of patients receiving combined anakinra and methotrexate treatment. Ann Rheum Dis 2005;64:1503–6.
- 20 Maksymowych WP, Landewé R, Conner-Spady B, Dougados M, Mielants H, Poole RA, et al. Serum matrix metalloproteinase 3 is an independent predictor of structural damage progression in patients with ankylosing spondylitis (AS). Arthritis Rheum, 2007. In press.
- 21 Davis J, van der Heijde D, Braun J, Dougados M, Cush J, Clegg D, et al. Recombinant human tumor necrosis factor receptor (etanercept) for treating ankylosing spondylitis. A randomized, controlled trial. Arthritis Rheum 2003;48:3230–6.
- 22 Braun J, Landewé R, Hermann KG, Han J, Yan S, Williamson P, et al. Major reduction in spinal inflammation in patients with ankylosing spondylitis after treatment with infliximab: results of a multicenter, randomized, double-blind, placebocontrolled magnetic resonance imaging study. Arthritis Rheum 2006;54:1646–52.
- controlled magnetic resonance imaging study. Arthritis Rheum 2006;54:1646–52.
 van der Heijde D, Kivitz A, Schiff M, Sieper J, Dijkmans B, Braun J, et al. Efficacy and safety of adalimumab in patients with ankylosing spondylitis: results of a multicenter, randomized, double-blind, placebo-controlled trial. Arthritis Rheum 2006;54:2136–2.
- 24 van der Heijde DM, Landewé RB, Ory P, Vosse D, Zhou L, Tsuji W, et al. Two-year etanercept therapy does not inhibit radiographic progression in patients with ankylosing spondylitis. Ann Rheum Dis 2006;65(Suppl II):81.